

# UC San Diego

## UC San Diego Previously Published Works

### Title

Cytokine and Lipid Mediator Regulation of Group 2 Innate Lymphoid Cells (ILC2s) in Human Allergic Airway Disease.

### Permalink

<https://escholarship.org/uc/item/3qp3670r>

### Journal

Journal of cytokine biology, 2(2)

### ISSN

2576-3881

### Authors

Cavagnero, Kellen  
Doherty, Taylor A

### Publication Date

2017-08-01

### DOI

10.4172/2576-3881.1000116

Peer reviewed



Published in final edited form as:

*J Cytokine Biol.* 2017 August ; 2(2): .

## Cytokine and Lipid Mediator Regulation of Group 2 Innate Lymphoid Cells (ILC2s) in Human Allergic Airway Disease

Kellen Cavagnero and Taylor A. Doherty\*

Department of Medicine, University of California San Diego, USA

### Abstract

The recent discovery of group 2 innate lymphoid cells (ILC2s) has caused a paradigm shift in the understanding of allergic airway disease pathogenesis. Prior to the discovery of ILC2s, Th2 cells were largely thought to be the primary source of type 2 cytokines; however, activated ILC2s have since been shown to contribute significantly, and in some cases, dominantly to type 2 cytokine production. Since the discovery of ILC2s in 2010, many mediators have been shown to regulate their effector functions. Initial studies identified the epithelial derived cytokines IL-25, IL-33, and TSLP as activators of ILC2s, and recent studies have identified many additional cytokine and lipid mediators that are involved in ILC2 regulation. ILC2s and their mediators represent novel therapeutic targets for allergic airway diseases and intensive investigation is underway to better understand ILC2 biology and upstream and downstream pathways that lead to ILC2-driven airway pathology. In this review, we will focus on the cytokine and lipid mediators that regulate ILC2s in human allergic airway disease, as well as highlight newly discovered mediators of mouse ILC2s that may eventually translate to humans.

### Keywords

Group 2 innate lymphoid cells (ILC2); Type 2 cytokine; Leukotrienes; Asthma; Rhinitis; IL-33; TSLP; Lipid mediators; Allergic airway disease; Aspirin Exacerbated Respiratory Disease (AERD)

### Introduction

Group 2 innate lymphoid cells (ILC2s) are a relatively newly discovered lymphocyte population that promotes features of allergic airway diseases including asthma and chronic rhinosinusitis through secretion of the type 2 cytokines IL-4, IL-5 and IL-13 among others. Unlike Th2 cells, ILC2s lack T cell receptors and, therefore, are not controlled in an antigen-specific manner [1]. Additionally, ILC2s lack surface receptors expressed by the major hematopoietic lymphocyte lineages including B, T, natural killer T (NKT), and natural killer (NK) cells [1,2]. In mouse models of asthma, ILC2s have been shown to drive airway

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

\*Corresponding author: Taylor A. Doherty, Department of Medicine, University of California San Diego, Biomedical Sciences Building, Room 5090, 9500 Gilman Drive, La Jolla, CA 92093-0635, USA, Tel: (858) 822-7653; Fax: (858) 534-2110; tdoherty@ucsd.edu.

eosinophilia, hyperresponsiveness, and mucous hypersecretion that are all features of allergic asthma [1,2]. Recently, increased levels of ILC2s have been detected in patients with allergic airway diseases including asthma, allergic rhinitis (AR), and chronic rhinosinusitis (CRS) [3–12]. Further, ILC2s are recruited to nasal mucosa from the blood in aspirin exacerbated respiratory disease (AERD), which is characterized by intense eosinophilic nasal polyps, asthma and respiratory reactions after COX-1 inhibition [13,14].

Studies examining the regulation of ILC2 homeostasis, activation, and proliferation have identified a myriad of mediators that both promote and inhibit their function. Early ILC2 studies using mouse models of asthma with protease-derived allergens demonstrated that damaged airway epithelial cells release cytokines that include IL-25, IL-33, and thymic stromal lymphopoietin (TSLP), which directly and potently activate ILC2s, leading to production of IL-4, IL-5, IL-9, and IL-13 [1,2]. Interestingly, certain ILC2 activators are capable of inducing select type 2 cytokine production. For example, stimulation of ILC2s with IL-33 results in IL-5 and IL-13 – yet minimal IL-4 – production, whereas stimulation with lipids, such as cysteinyl leukotrienes (CysLTs) and prostaglandin D2, causes robust IL-4 production [15]. Furthermore, combinations of ILC2 activators that utilize non-redundant signaling pathways can result in additive and even synergistic activation [16].

Each Th2 cytokine downstream of ILC2s plays an important role in disease pathogenesis: IL-4 drives naïve T cell differentiation into Th2 cells and B cell IgE class switching, and supports ILC2 homeostasis; IL-5 drives airway eosinophilia; IL-9 stimulates ILC2 cell proliferation and inhibits apoptosis in an autocrine fashion; and IL-13 induces airway hyper responsiveness, goblet cell metaplasia, mucous hyper secretion, and IgE class switching [1,2]. This review will largely focus on the upstream cytokine and lipid mediators that regulate ILC2s in human allergic airway disease, which are summarized in Figure 1. In addition, we will highlight newly discovered mediators of mouse ILC2s that may also translate to humans.

## Epithelial Cytokines that Regulate ILC2s

### IL-33

IL-33 has been shown to be a potent driver of experimental allergic airway disease in mice, and its expression has been linked to many human allergic airway diseases [17]. In humans, IL-33 expression is highly upregulated in AERD patient nasal polyps, AR patient nasal secretions, and asthmatic airways (with severe asthmatics demonstrating the highest expression) [14,18,19]. Surprisingly, IL-33 expression is not upregulated in CRS polyps, though the IL-33 receptor has been detected at higher levels [19]. Importantly, large population genome wide association studies have shown that small nucleotide polymorphisms (SNPs) involving IL33 and its receptor are associated with increased asthma susceptibility [20].

IL-33 is classically considered an epithelial derived cytokine; however, endothelial cells, smooth muscle cells, and macrophages also express IL-33 [1,2,18]. IL-33 and IL-25, which is also epithelial cell-derived, were the first cytokines demonstrated to activate murine ILC2s [21]. In mice, IL-33 administration induces a robust type 2 immune response independent of

adaptive immunity, which was one of the first clues to the presence of a novel non-T cell producing Th2 cytokines [2,22]. The IL-33 receptor is a heterodimeric receptor composed of ST2 and IL-1R (T1/ST2) [2]. Additional cell types including Th2 cells, eosinophils, dendritic cells (DCs), macrophages, basophils, mast cells, and NKT cells also express T1/ST2 and are activated by IL-33 [1,2]. In a landmark 2011 human study, ILC2s in peripheral blood and fetal gut were shown to produce IL-13 in response to IL-33 stimulation *in vitro* [9]. Importantly, the authors were the first to identify ILC2s in human lung tissue [9]. Overall, these findings suggest that the diseased human airway is an environment where IL-33 and/or its receptor are increased along with mucosal ILC2s. Thus, IL-33-induced activation of ILC2s very likely plays a role in human allergic airway disease pathogenesis.

## IL-25

Similar to IL-33, the epithelial cytokine IL-25 has been shown to promote experimental allergic airway disease in mice, and its expression has been linked to human allergic airway disease. Corrigan et al. found that human asthmatic bronchial biopsies from patients challenged with allergen had increased numbers of IL-25+ and IL-25R+ cells, and Lee et al. found increased IL-25 expression in CRS patient nasal polyp tissue [23,24]. To our knowledge, no studies have found a correlation between IL-25, and/or its receptor, and AR or AERD patient tissue.

Similar to IL-33, IL-25 is classically considered to be an epithelial cytokine; however, endothelial cells, eosinophils, Th2 cells, and mast cells have also been shown to secrete IL-25 [1,2,23]. IL-25 binds to the IL-25R receptor—a heterodimeric receptor consisting of IL-17RA and IL-17RB—which is expressed on the surface of ILC2s, Th2 cells, eosinophils, DCs, macrophages, and NKT cells [2]. The initial human ILC2 study that showed that IL-33 activates ILC2s also found that ILC2s in peripheral blood and fetal gut produce IL-13 in response to IL-25 stimulation [9]. Though IL-25 appears to serve a similar function to IL-33, experimental mouse models of asthma have elucidated some important differences. Mice demonstrate peak expression of IL-33 three hours post-allergen challenge, whereas IL-25 peaks, but to a lesser magnitude than IL-33, twelve hours post-challenge [25]. Additionally, IL-33 has been shown to be more potent than IL-25 in mouse models though relevance to humans is not clear [25]. Thus, because IL-25-induced activation of ILC2s may contribute to human asthma and CRSwNP pathogenesis, IL-25 represents a potential therapeutic target.

## Thymic Stromal Lymphopoietin (TSLP)

Thymic stromal lymphopoietin (TSLP) is an epithelial cytokine that was detected at increased levels in allergic airway disease well before ILC2s were discovered [26]. TSLP expression is increased in nasal polyps from CRS and AERD patients, AR patient nasal tissue, and in asthmatic airway epithelial cells (with levels positively correlating with disease severity) [14,27–29]. Additionally, TSLP SNPs that result in increased TSLP activity are associated with increased susceptibility to asthma [30].

Initially, TSLP was shown to prime DCs for adaptive Th2 responses and this role was thought to be primarily responsible for downstream type 2 inflammation driven by TSLP [26]. In addition to responding to TSLP, a recent human study showed that dendritic cells

exposed to fungal allergen are also a potent source of TSLP [31]. Furthermore, recent studies have identified basophils and mast cells as additional sources of TSLP [32]. In humans, TSLP exists in two isoforms, long and short, which initiate inflammatory and homeostatic pathways, respectively [32]. Poposki et al. showed that TSLP fragments more potently activate type 2 immune responses compared to the full-length isoform [33]. TSLP exerts its effects by binding to the TSLPR and IL-7Ra heterodimeric receptor, which is expressed on ILC2s, DCs, and mast cells [33,34].

Aside from its known role in adaptive immunity, TSLP has been found to be a potent activator of ILC2s, independent of IL-33 [35]. In a landmark study, Mjosberg et al. showed that TSLP-stimulated ILC2s from peripheral blood and nasal polyp tissue upregulated GATA3, the master transcriptional regulator of type 2 cytokines, and produced IL-4, IL-5, IL-13 [36]. The effect of TSLP-driven activation of ILC2s was enhanced by addition of IL-33. Interestingly, a recent study by Liu et al. found that, of the epithelial cell-derived cytokines, only TSLP was able to induce ILC2 corticosteroid resistance (an effect also known to be present in mouse ILC2s) [37,38]. Importantly, an anti-TSLP antibody was also recently shown to significantly reduce airway eosinophils and hyperresponsiveness in asthmatics following allergen challenge [39]. Together, these results suggest that TSLP is a potent inducer of ILC2-driven type 2 inflammation as well as adaptive Th2 responses in humans; therefore, TSLP represents a therapeutic target for allergic airway diseases.

## Additional Cytokine Regulation of ILC2s

### IL-2 and IL-7

The development of ILC2s from common lymphoid progenitor cells requires IL-7R and IL-2R expression, and mice lacking IL-2R or IL-7R lack functional ILC2s [21,40]. IL-7 is a hematopoietic growth factor produced by epithelial cells and DCs that binds to IL-7R on ILC2s to promote homeostasis and proliferation [41]. IL-2 is produced by many lymphocytes and also promotes ILC2 homeostasis and proliferation by acting through the IL-2 receptor [41]. Though IL-2 and IL-7 are largely homeostatic for ILC2 responses, there is one report that demonstrated that IL-2 treatment of RAG1 deficient mice (lack B and T cells, but have ILC2s) led to increased ILC2 proliferation, activation, and dermatitis [42]. This suggests that, under specific conditions, IL-2 could be pro-inflammatory through ILC2 activation.

### IL-1 and IL-12

In 2016, two independent groups reported that IL-1 cytokines were capable of activating ILC2s and promoting ILC plasticity [43,44]. *In vitro*, IL-1 $\alpha$  and IL-1 $\beta$  were found to potently induce human peripheral blood ILC2 proliferation and IL-5 and IL-13 production. Interestingly, IL-1 $\beta$  caused ILC2s to upregulate epithelial cell-derived cytokine receptors, while also priming ILC2s for conversion into ILC1s. These findings were confirmed *in vivo*, where the presence of IL-12 was the determining factor of whether IL-1 $\beta$  stimulation would result in ILC2 activation or ILC2 plasticity. The presence of IL-12 reduced ILC2 GATA3 expression and shifted IL-1 $\beta$  from an ILC2-activating cytokine to an ILC1-inducing cytokine.

## IL-4

IL-4 expression is increased in AERD, asthma, CRSwNP, CRSsNP, and AR [14,45,46]. Additionally, IL-4 SNPs are associated with increased asthma and AR risk [46]. ILC2s, mast cells, eosinophils, and basophils are capable of producing IL-4, and ILC2s also express the IL-4 receptor [2,43,47]. Thus, IL-4 can act in an autocrine fashion on ILC2s. Initial animal studies examining the relationship between IL-4 and ILC2s found that mouse lung ILC2s upregulated IL-9, IL-13, CCL3, CCL5, and CCL11 expression following IL-4 stimulation *in vitro*, and IL-33 potentiated this activation [48]. Interestingly, IL-5 was not upregulated with IL-4 alone, which suggests distinct activation mechanisms. Further, IL-4 alone was not sufficient to induce proliferation, but it did enhance IL-33-driven ILC2 proliferation. A recent human study found IL-4 to play a role similar to IL-2 and IL-7 in ILC2 maintenance [43]. Like prior animal studies, IL-4 was shown to potentiate IL-33-driven ILC2 proliferation and cytokine production; however, in contrast to the previous animal studies, IL-4 alone was not sufficient to induce cytokine production. Importantly, IL-4 was found to upregulate the expression of the PGD2 receptor CRTH2, which could then render ILC2s more responsive to PGD2. Furthermore, IL-4 prevented ILC2 to ILC1 conversion. Thus, IL-4 plays a unique role in ILC2 maintenance, proliferation, and activation. Multiple IL-4-inhibiting monoclonal antibody therapies have been tested in asthma clinical trials though the antibodies largely did not display efficacy. However, targeting IL-4R/IL-13R with dupilumab was recently FDA approved for asthma based on strong clinical trial data [49,50].

## TL1A

TL1A, a member of the TNF superfamily, has recently been shown to regulate mouse and human ILC2s. Activated T cells, myeloid cells, and endothelial cells are known sources of TL1A [51]. TL1A is the ligand for the DR3 receptor, which is expressed by ILC2s, T cells, and NKT cells [1,51,52]. Two recent studies reported that TL1A potentially activates ILC2s [51,52]. Meylan et al. showed that mouse mesenteric lymph node (mLN) ILC2s stimulated *in vitro* with TL1A alone induced IL-5, IL-6, IL-13, and IL-17A secretion, and this effect was potentiated by IL-7. Interestingly, TL1A only induced IL-9 production when combined with IL-7. In contrast to their *in vitro* findings, exogenous TL1A was found to have no effect on ILC2 proliferation or cytokine production *in vivo*. Yu et al. confirmed Meylan et al.'s *in vitro* findings that murine mLN ILC2s stimulated with TL1A alone produce significantly more IL-5 and IL-13 compared to control. However, Yu et al.'s work showed that exogenous TL1A administration was sufficient to increase mouse ILC2 numbers and cytokine production *in vivo*. Additionally, the authors showed that TL1A alone was sufficient to induce human ILC2 IL-5 and IL-13 production and further potentiated IL-25 and IL-33-induced ILC2 cytokine production. Taken together, these findings suggest that TL1A could play a role in human allergic airway disease and represents a novel potential target for therapeutic intervention.

## IL-9

In 2011, Wilhelm et al. was the first to report that IL-9 maintains activated ILC2s through IL-9R [53]. In 2013, using a mouse helminth model of type 2 inflammation, the same group demonstrated that IL-9 potentiates ILC2 IL-5 and IL-13 production in an autocrine fashion

[54]. Unlike IL-7, IL-9 was able to effectively maintain only activated ILC2s, rather than all ILC2s, and protect them from activation-induced apoptosis. Thus, IL-9 could be critical to sustaining type 2 inflammation through ongoing ILC2 activation. Additional studies are necessary to determine whether IL-9 has similar effects on ILC2 function in humans.

### Interferons & IL-27

A recent animal study reported that the Th1 cytokines IFN- $\gamma$  and IL-27, and the type 1 interferon IFN- $\beta$ , potently reduce ILC2 proliferation and cytokine production by directly targeting ILC2s [55]. Interestingly, the authors found that the ability of these cytokines to suppress ILC2 function was dependent upon localization: ILC2s from the mLN were sensitive to IL-27 suppression and resistant to IFN- $\gamma$ , whereas tissue resident ILC2s were more sensitive to IFN- $\gamma$ .

Furthermore, a recent report showed that IFN- $\beta$  potently and directly suppressed mouse lung ILC2 proliferation and IL-5 production [56]. The group also demonstrated that purified human cord blood-derived ILC2s exposed to IFN- $\beta$  had significantly reduced cytokine production and proliferation. Importantly, IFN SNPs are associated with asthma, and treatment of asthmatics with IFNs has been shown to reduce type 2 airway pathology.

### IL-10 and TGF- $\beta$

ILC2s highly express IL-10R and TGF- $\beta$  receptors, and regulatory T cell (Treg) IL-10 and TGF- $\beta$  production has recently been reported to directly inhibit IL-33-induced ILC2 IL-5 and IL-13 production [57,58]. Interestingly, the costimulatory ligand/receptor pair ICOS/ICOSL was required for full Treg mediated suppression, suggesting both direct (cell contact) and indirect (cytokines) dampening of ILC2s by Tregs. Additional studies are required to determine whether targeting Tregs during a natural Th2 disease course has an effect on ILC2s.

### Lipid Mediators that Regulate ILC2s

A plethora of cell types produce eicosanoids that, in turn, act on many different cell types to drive inflammation. In allergic airway disease pathophysiology, the primary producers of eicosanoids are activated mast cells, eosinophils, DCs, and macrophages [59]. The eicosanoid family members, prostaglandins (PGs) and leukotrienes (LTs), are bioactive lipid products of arachidonic acid (AA) metabolism. Two distinct metabolic pathways downstream of AA are the cyclooxygenase (COX) pathway, which produces PGs, and the 5-lipoxygenase (5LO) pathway, which produces LTs.

### Leukotrienes

The 5LO pathway of AA metabolism first produces LTA<sub>4</sub>, which can then be converted to the parent cysteinyl leukotriene (CysLT) LTC<sub>4</sub> as well as LTB<sub>4</sub>. LTB<sub>4</sub> is a final metabolite in its pathway, whereas LTC<sub>4</sub> is rapidly converted into LTD<sub>4</sub>, and then into the terminal CysLT LTE<sub>4</sub> [59].



## Cysteinyl Leukotrienes (CysLTs)

The CysLTs LTC<sub>4</sub>, LTD<sub>4</sub>, and LTE<sub>4</sub> are elevated in asthma, allergic rhinitis, AERD, and CRS [59]. CysLTs were initially identified as a cause of bronchoconstriction after CysLT aerosol administration resulted in airway hyper responsiveness in both asthmatics and non-asthmatics [60]. Since then, several reports have established CysLTs in pro-inflammatory roles during type 2 immunity [61–63]. There are two primary CysLT receptors, CysLT<sub>1</sub>R and CysLT<sub>2</sub>R. CysLT<sub>1</sub>R and CysLT<sub>2</sub>R are capable of binding all of the CysLTs; however, CysLT<sub>1</sub>R binds LTD<sub>4</sub> with the greatest affinity and has a low affinity for LTE<sub>4</sub>, while CysLT<sub>2</sub>R binds LTD<sub>4</sub> and LTC<sub>4</sub> with equal affinity and also has a low affinity for LTE<sub>4</sub> [16]. Recently, additional receptors, P2Y<sub>12</sub> and GPR99, were shown to mediate responses to LTE<sub>4</sub> [64].

In 2013, Doherty et al. was the first to report that mouse lung ILC2s express CysLT<sub>1</sub>R and ILC2s are activated *in vitro* and *in vivo* by LTD<sub>4</sub> [15]. CysLT-induced ILC2 activation and calcium influx was found to be dependent on CysLT<sub>1</sub>R as activation was abrogated by administration of montelukast, a CysLT<sub>1</sub>R antagonist. Interestingly, ILC2 activation driven by the fungal allergen *Alternaria* was potentiated by LTD<sub>4</sub>, which suggests IL-33 and LTD<sub>4</sub> use non-redundant signaling mechanisms as *Alternaria* primarily mediates ILC2 responses through IL-33 [65]. Very recently, two studies confirmed that mouse lung ILC2s are activated, both *in vitro* and *in vivo*, by all of the CysLTs and that IL-33-induced ILC2 activation is further potentiated by CysLTs *in vitro* and *in vivo* [16,66].

The effect of CysLTs on human ILC2s appears to be conserved as Salimi et al. recently reported that: human ILC2s express CysLT<sub>1</sub>R, ILC2s are activated by CysLTs, and LTE<sub>4</sub> potentiates IL-33 and IL-25 induced ILC2 activation in a CysLT<sub>1</sub>R-dependent manner [67,68]. Interestingly, a previous report showed that LTE<sub>4</sub>-induced ILC2 IL-5 production *in vivo* was not blocked by montelukast, though this could be due to differences between humans and mice and/or conditions (*in vivo* vs. *in vitro*) [15]. Overall, CysLTs are potent activators of ILC2s, in both humans and mice, and are largely CysLT<sub>1</sub>R-dependent. While CysLT<sub>1</sub>R inhibition has long been used in the clinic to manage asthma, AR, CRS, and AERD symptoms, it is possible that combinational therapies blocking CysLTs and epithelial cell-derived cytokines will further alleviate type 2 cytokine-driven symptoms. Additional studies are needed to elucidate the function of the CysLT<sub>2</sub>, GPR99, and P2Y<sub>12</sub> receptors in ILC2-driven allergic airway inflammation.

## Leukotriene B<sub>4</sub> (LTB<sub>4</sub>)

LTB<sub>4</sub> binds the LTB<sub>4</sub>R<sub>1</sub> and LTB<sub>4</sub>R<sub>2</sub> receptors. LTB<sub>4</sub>R<sub>1</sub> is the primary receptor for LTB<sub>4</sub> and is known to mediate chemotaxis whereas LTB<sub>4</sub>R<sub>2</sub> binds LTB<sub>4</sub>, and other eicosanoids, with a relatively low affinity [69]. Recently, Moltke et al. reported high levels of LTB<sub>4</sub>R<sub>1</sub>, and not LTB<sub>4</sub>R<sub>2</sub>, expression on mouse lung ILC2s [16]. The LTB<sub>4</sub>R<sub>1</sub> receptor was found to be functional as LTB<sub>4</sub> stimulation *in vitro* increased ILC2 IL-13 production and was ameliorated in ILC2s lacking the LTB<sub>4</sub>R<sub>1</sub> receptor. Interestingly, no effect of LTB<sub>4</sub> was found *in vivo*. Additional studies are necessary to elucidate the role of LTB<sub>4</sub> in human ILC2-driven allergic airway disease.



## Prostaglandins

The COX pathway of AA metabolism generates a wide variety of PGs. However, the PGs that will be discussed in this review are those that have been shown to regulate ILC2s, PGD2 and PGI2.

### Prostaglandin D2 (PGD2)

PGD2 expression is elevated in patients with severe asthma, CRS, AR, and is a dominant contributor to AERD pathology [14,70]. PGD2 is primarily synthesized by mast cells, but can also be produced by eosinophils [71]. Binding of PGD2 to CRTH2 expressed by ILC2s, Th2 cells, eosinophils, and basophils and results in type 2 cytokine production, chemotaxis, and increased cell survival [71].

In 2011, human ILC2s were first identified using the CRTH2 receptor, which distinguishes ILC2s from ILC1s and ILC3s [9]. A later study showed that human peripheral blood ILC2s produce increased IL-13 in response to PGD2 in the presence of IL-2, IL-25, and IL-33 [72]. PGD2 was subsequently found to induce ILC2 chemotaxis, an effect that was blocked by a CRTH2 inhibitor [71]. Xue et al. confirmed that PGD2 activates and induces chemotaxis of human ILC2s as well as potentiates the effect of IL-25 and IL-33 in the presence of IL-2 [67]. Aside from the classic ILC2 cytokines IL-4, IL-5, IL-13, the group also found additional pro-inflammatory cytokines produced by ILC2s including IL-3, IL-8, IL-9, IL-21, GM-CSF, and CSF1. Further, they showed that ILC2s up-regulated IL-25 and IL-33 receptors following PGD2 stimulation and hypothesized that PGD2 primed ILC2s for a potentiating effect by IL-25 and IL-33. In 2015, Wojno et al. showed that human ILC2 CRTH2 expression is tissue specific: peripheral blood ILC2s express higher levels of CRTH2 than lung ILC2s. Further, the group used a mouse model to show that CRTH2 regulates ILC2 chemotaxis *in vivo* [73]. The reduction of CRTH2 on human tissue ILC2s is an important finding as human ILC2s are identified by their CRTH2 expression. Therefore, studies of human tissue ILC2s may be excluding CRTH2-negative or -low populations when identifying ILC2s. Thus, it is vital that unique and constitutive identifying markers for ILC2s are discovered. Overall, these findings strongly demonstrate that PGD2 has multiple effects on ILC2s that could contribute to disease pathogenesis.

### Prostaglandin I2 (PGI2)

PGI2, another PG product of AA metabolism, has also been implicated in human allergic airway disease. Genome wide association studies previously showed that PGI2 SNPs are associated with aspirin intolerant asthma [74], and the PGI2 metabolite, 6-keto-PGF1 $\alpha$ , was elevated in human lung tissue during *in vitro* anaphylaxis assays [75]. In the clinic, PGI2 analogs are commonly used to treat pulmonary hypertension; however, studies examining the therapeutic effect of PGI2 in human allergic airway disease have, thus far, been inconclusive. One study showed no change in FEV1 (expired lung volume in 1 second) and airway hyper responsiveness in response to methacholine challenge after PGI2 administration, while another study saw a decrease in FEV1 [76,77].

PGI2 is highly expressed in lung tissue and is produced by smooth muscle cells, endothelial cells, fibroblasts, follicular dendritic cells, and thymic nurse cells [76,78]. PGI2 has predominately anti-inflammatory properties and exerts its effects through binding of the prostacyclin receptor (IP), which is expressed by many immune cell types including bone marrow-derived DCs, Th1 cells, Th2 cells, and ILC2s [77]. PGI2-IP interactions ultimately result in smooth muscle relaxation, reduced cellular proliferation, and anti-inflammatory mechanisms including increased secretion of IL-10 by Tregs [78].

In 2016, Zhou et al. was the first to report that PGI2 inhibits ILC2 function [79]. In this study, bone marrow-derived ILC2s stimulated with IL-33 *in vitro* had reduced IL-5 and IL-13 production and proliferation in the presence of Cicaprost, a PGI2 analog. This attenuation was prevented in ILC2s lacking the IP receptor. Further, *Alternaria*-challenged IP knockout mice exhibited both increased numbers of lung ILC2s and increased ILC2 IL-5 and IL-13 production compared to wild type mice. Furthermore, wild type mice given both Cicaprost and *Alternaria* had lower levels of ILC2s and type 2 cytokine production, compared to mice given *Alternaria* alone. Finally, Cicaprost was found to inhibit IL-33-stimulated human peripheral blood ILC2 IL-5 and IL-13 production. These findings suggest that PGI2 might inhibit ILC2-driven allergic airway inflammation in humans.

## Specialized Pro-Resolving Lipid Mediators

Lipoxin A4 (LXA4) is generated by the 15-lipoxygenase (15LO) pathway in eosinophils, epithelial cells, and macrophages. Patients with severe asthma have reduced levels of LXA4 and its receptor, ALX/FPR2. A 1992 study found that patients given LXA4 following LTC4 challenge exhibited a reduction of asthmatic airway responses suggesting a protective role of LXA4 in asthma [70,80]. Recently, Barnig et al. found that human peripheral blood ILC2s express the LXA4 receptor and that LXA4 inhibited ILC2 responses in the presence of IL-25, IL-33, and PGD2 *in vitro* [72].

Maresin-1 is a product of omega-3 fatty acid metabolism. One report investigated the effects of maresin-1 and ILC2 responses [81]. Using a metabolipidomics approach, Krishnamoorthy et al. found that maresin-1 is increased in mouse lung tissue during the resolution phase of allergic inflammation. Maresin-1 administration *in vivo* inhibited ILC2 activation and promoted the generation of Tregs resulting in the resolution of inflammation. Additional studies are necessary to determine the role of maresin-1 in human ILC2 responses.

## ILC2 Regulation in Human Airway Disease

### Asthma

Asthma is a chronic lung disease characterized by airway inflammation, increased mucous production, hyperresponsiveness and remodeling, along with symptoms that include dyspnea, wheezing, chest tightness, and coughing [1,2]. Although there are many subtypes of asthma, a large portion of asthmatics have high levels of type 2 cytokines in their airways [82]. This suggests contributions from ILC2s in addition to CD4<sup>+</sup> Th2 cells. Several pro-inflammatory cytokines (IL-25, IL-33 and TSLP) and lipid (CysLTs and PGD2) mediators are also elevated in asthmatic airways, which could drive ILC2 responses [18,23,27,59,70].

In 2013, the first report examining ILC2s and human asthma found no difference in peripheral blood ILC2 numbers between severe asthmatics, mild asthmatics, and healthy controls [72]. However, subsequent reports have shown differences in ILC2 numbers and activation between asthmatics and non-asthmatics. In 2014, Bartemes et al. found increased numbers of peripheral blood ILC2s and Th2 cytokine responses in allergic asthmatics compared to controls [3]. This finding was supported by a 2016 study that found increased numbers of activated ILC2s in the blood and sputum of severe asthmatics compared to mild, atopic asthmatics and healthy controls [4]. Furthermore, increased numbers of ILC2s have been found in both adult and pediatric asthma patients compared to controls [5,6]. Very recent studies have also demonstrated that ILC2s are increased after allergen challenge in asthmatics and that TSLP imparts ILC2 corticosteroid resistance in severe asthmatics [37,83]. The precise role of ILC2s in human asthma remains to be elucidated; however, these findings along with a multitude of animal studies suggest that ILC2s may contribute significantly to disease pathogenesis.

### Allergic rhinitis

Allergic rhinitis is caused by an IgE-mediated and type 2 inflammatory response to inhaled allergens which results in nasal symptoms including sneezing, itchiness, difficulty breathing, and discharge [84]. Previous work demonstrated that challenging sensitized AR patients with allergens causes a rapid increase in PGD<sub>2</sub> production. Given that PGD<sub>2</sub> has been shown to cause ILC2 activation, proliferation, and chemotaxis, it is plausible that ILC2s play a role in AR pathogenesis. In support of ILC2 activation in AR, IL-33 expression was found to be increased in the nasal secretions of patients with AR compared to control patients [85].

The first study to examine the relationship between AR and ILC2s found that nasal cat allergen challenge of cat-sensitized AR patients led to significantly increased percentage of ILC2s in peripheral blood compared to control challenge [7]. Consistent with this finding, a subsequent report found that peripheral blood ILC2s were increased in patients with grass pollen-sensitized AR during the pollen season compared to control patients, and that ILC2 levels were reduced by subcutaneous immunotherapy [8]. However, a separate report found that patients with AR have neither enhanced type 2 responses nor increased levels of ILC2s in the peripheral blood [3]. A recent study may shed light on these differences in results as subtypes of AR including House dust mite (HDM)- and mugwort-mediated AR were found to have differences in ILC2 responses [86]. The authors showed that peripheral blood ILC2s were increased, and had greater activation status, in patients with HDM-AR compared to mugwort-AR. The authors speculated that the differences could be due to the different mechanisms of action and/or levels of immunogenicity. Taken together, these findings demonstrate that ILC2s may promote specific subtypes of AR. Additional studies are needed to illuminate the overall role of ILC2s in this IgE driven nasal disorder.

### Chronic rhinosinusitis

Chronic rhinosinusitis is an inflammatory disease of the nasal and paranasal sinuses and is often associated with asthma and AR [2]. In some cases, CRS pathogenesis includes the development of nasal polyps (CRSwNP), which are classified as eosinophilic or non-eosinophilic. Nasal polyps from patients with CRS have elevated levels of CysLTs, IL-33,

TSLP, and IL-4 [2,34] and multiple groups have shown that ILC2s are enriched in nasal polyps, especially eosinophilic polyps, compared to control tissue [9–12]. Together, these findings suggest that ILC2s may contribute to CRS pathogenesis and that eosinophilic polyposis is ILC2-driven.

## Aspirin Exacerbated Respiratory Disease (AERD)

AERD is a chronic inflammatory disease of the respiratory tract that includes moderate to severe asthma, nasal polyps, and respiratory reactions to COX-1 enzyme inhibitor medications [14]. Respiratory tissue from AERD patients has intense tissue eosinophilia, mast cell activation, and high levels of PGs (specifically PGD<sub>2</sub>) and LTs [14,87]. Recently, elevated levels of IL-4, TSLP, and IL-33 have also been found in patients with AERD [14,88,89]. The findings of increased PGD<sub>2</sub>, CysLTs, IL-33, and TSLP in AERD, as well as eosinophilic nasal polyps enriched in ILC2s, strongly suggest that ILC2s play a role in AERD pathogenesis. In 2017, Eastman et al. were the first to identify changes in ILC2s in AERD. Increased numbers of ILC2s were found in the nasal mucosa, and decreased numbers of ILC2s in the peripheral blood, during aspirin (COX-1 inhibitor) challenge in AERD patients [13]. One possible explanation for this is that ILC2s migrate from the peripheral blood to the nasal mucosa during reactions in response to PGD<sub>2</sub> interacting with CRTH2 on ILC2s. Additional studies are necessary to elucidate the functional role, and possible mechanism of migration, of ILC2s in AERD.

## Summary

In the last decade since ILC2s have been discovered, a large number of cytokines and lipid mediators have been shown to positively and negatively regulate ILC2 homeostasis, proliferation, and activation in both mouse models of airway disease and in human samples. ILC2s and many positive regulators including CysLTs, PGD<sub>2</sub>, IL-33 and TSLP are elevated in the airways of patients with allergic diseases, which supports an environment of ILC2-driven inflammation. Development of therapies that target specific mediators upstream of ILC2s are underway and the challenge of uncovering ILC2-specific therapy remains. However, based on the discovery of negative regulators of ILC2s and finding that combinations of specific mediators can potentiate ILC2-driven inflammation, it is plausible that combinational therapies could better treat allergic airway disease and improve the quality of life of patients.

## Acknowledgments

T.A.D. is supported by NIH AI114585 and AI070535.

## References

1. Karta MR, Broide DH, Doherty TA. Insights into Group 2 Innate Lymphoid Cells in Human Airway Disease. *Curr Allergy Asthma Rep.* 2016; 16:8. [PubMed: 26746844]
2. Doherty TA, Broide DH. Group 2 innate lymphoid cells: new players in human allergic diseases. *J Investig Allergol Clin Immunol.* 2015; 25:1–11.

3. Bartemes KR, Kephart GM, Fox SJ, Kita H. Enhanced innate type 2 immune response in peripheral blood from patients with asthma. *J Allergy Clin Immunol.* 2014; 134:671–678. [PubMed: 25171868]
4. Smith SG, Chen R, Kjarsgaard M, Huang C, Oliveria JP, et al. Increased numbers of activated group 2 innate lymphoid cells in the airways of patients with severe asthma and persistent airway eosinophilia. *J Allergy Clin Immunol.* 2016; 137:75–86. e8. [PubMed: 26194544]
5. Christianson CA, Goplen NP, Zafar I, Irvin C, Good JT Jr, et al. Persistence of asthma requires multiple feedback circuits involving type 2 innate lymphoid cells and IL-33. *J Allergy Clin Immunol.* 2015; 136:59–68. e14. [PubMed: 25617223]
6. Nagakumar P, Denney L, Fleming L, Bush A, Lloyd CM, et al. Type 2 innate lymphoid cells in induced sputum from children with severe asthma. *J Allergy Clin Immunol.* 2016; 137:624–626. [PubMed: 26277593]
7. Doherty TA, Scott D, Walford HH, Khorram N, Lund S, et al. Allergen challenge in allergic rhinitis rapidly induces increased peripheral blood type 2 innate lymphoid cells that express CD84. *J Allergy Clin Immunol.* 2014; 133:1203–5. [PubMed: 24582313]
8. Lao-Araya M, Steveling E, Scadding GW, Durham SR, Shamji MH, et al. Seasonal increases in peripheral innate lymphoid type 2 cells are inhibited by subcutaneous grass pollen immunotherapy. *J Allergy Clin Immunol.* 2014; 134:1193–5. e4. [PubMed: 25212194]
9. Mjösberg JM, Trifari S, Crellin NK, Peters CP, van Drunen CM, et al. Human IL-25- and IL-33-responsive type 2 innate lymphoid cells are defined by expression of CCR2 and CD161. *Nat Immunol.* 2011; 12:1055–1062. [PubMed: 21909091]
10. Walford HH, Lund SJ, Baum RE, White AA, Bergeron CM, et al. Increased ILC2s in the eosinophilic nasal polyp endotype are associated with corticosteroid responsiveness. *Clin Immunol.* 2014; 155:126–135. [PubMed: 25236785]
11. Miljkovic D, Bassiouni A, Cooksley C, Ou J, Hauben E, et al. Association between group 2 innate lymphoid cells enrichment, nasal polyps and allergy in chronic rhinosinusitis. *Allergy.* 2014; 69:1154–1161. [PubMed: 24924975]
12. Ho J, Bailey M, Zaunders J, Mrad N, Sacks R, et al. Group 2 innate lymphoid cells (ILC2s) are increased in chronic rhinosinusitis with nasal polyps or eosinophilia. *Clin Exp Allergy.* 2015; 45:394–403. [PubMed: 25429730]
13. Eastman JJ, Cavagnero KJ, Deconde AS, Kim AS, Karta MR, et al. Group 2 innate lymphoid cells are recruited to the nasal mucosa in patients with aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol.* 2017; 140:101–108. [PubMed: 28279492]
14. Woessner KM. Update on Aspirin-Exacerbated Respiratory Disease. *Curr Allergy Asthma Rep.* 2017; 17:2. [PubMed: 28097500]
15. Doherty TA, Khorram N, Lund S, Mehta AK, Croft M, et al. Lung type 2 innate lymphoid cells express cysteinyl leukotriene receptor 1, which regulates TH2 cytokine production. *J Allergy Clin Immunol.* 2013; 132:205–213. [PubMed: 23688412]
16. Von Moltke J, O'Leary CE, Barrett NA, Kanaoka Y, Austen KF, et al. Leukotrienes provide an NFAT-dependent signal that synergizes with IL-33 to activate ILC2s. *J Exp Med.* 2017; 214:27–37. [PubMed: 28011865]
17. Drake LY, Kita H. IL-33: biological properties, functions, and roles in airway disease. *Immunol Rev.* 2017; 278:173–184. [PubMed: 28658560]
18. Préfontaine D, Lajoie-Kadoch S, Foley S, Audusseau S, Olivenstein R, et al. Increased expression of IL-33 in severe asthma: evidence of expression by airway smooth muscle cells. *J Immunol.* 2009; 183:5094–5103. [PubMed: 19801525]
19. Baba S, Kondo K, Kanaya K, Suzukawa K, Ushio M, et al. Expression of IL-33 and its receptor ST2 in chronic rhinosinusitis with nasal polyps. *Laryngoscope.* 2014; 124:E115–122. [PubMed: 24122812]
20. Moffatt MF, Gut IG, Demenais F, Strachan DP, Bouzigon E, et al. A large-scale, consortium-based genomewide association study of asthma. *N Engl J Med.* 2010; 363:1211–1221. [PubMed: 20860503]
21. Doherty TA. At the bench: understanding group 2 innate lymphoid cells in disease. *J Leukoc Biol.* 2015; 97:455–467. [PubMed: 25473099]

22. Kondo Y, Yoshimoto T, Yasuda K, Futatsugi-Yumikura S, Morimoto M, et al. Administration of IL-33 induces airway hyperresponsiveness and goblet cell hyperplasia in the lungs in the absence of adaptive immune system. *Int Immunol*. 2008; 20:791–800. [PubMed: 18448455]
23. Corrigan CJ, Wang W, Meng Q, Fang C, Eid G, et al. Allergen-induced expression of IL-25 and IL-25 receptor in atopic asthmatic airways and late-phase cutaneous responses. *J Allergy Clin Immunol*. 2011; 128:116–124. [PubMed: 21570719]
24. Lee M, Kim DW, Shin HW. Targeting IL-25 as a novel therapy in chronic rhinosinusitis with nasal polyps. *Curr Opin Allergy Clin Immunol*. 2017; 17:17–22. [PubMed: 27870664]
25. Barlow JL, Peel S, Fox J, Panova V, Hardman CS, et al. IL-33 is more potent than IL-25 in provoking IL-13-producing nuocytes (type 2 innate lymphoid cells) and airway contraction. *J Allergy Clin Immunol*. 2013; 132:933–941. [PubMed: 23810766]
26. Liu YJ. Thymic stromal lymphopoietin: master switch for allergic inflammation. *J Exp Med*. 2006; 203:269–273. [PubMed: 16432252]
27. Ying S, O'Connor B, Ratoff J, Meng Q, Mallett K, et al. Thymic stromal lymphopoietin expression is increased in asthmatic airways and correlates with expression of Th2-attracting chemokines and disease severity. *J Immunol*. 2005; 174:8183–8190. [PubMed: 15944327]
28. Kimura S, Pawankar R, Mori S, Nonaka M, Masuno S, et al. Increased expression and role of thymic stromal lymphopoietin in nasal polyposis. *Allergy Asthma Immunol Res*. 2011; 3:186–193. [PubMed: 21738884]
29. Cianferoni A, Spergel J. The importance of TSLP in allergic disease and its role as a potential therapeutic target. *Expert Rev Clin Immunol*. 2014; 10:1463–1474. [PubMed: 25340427]
30. Harada M, Hirota T, Jodo AI, Hitomi Y, Sakashita M, et al. Thymic stromal lymphopoietin gene promoter polymorphisms are associated with susceptibility to bronchial asthma. *Am J Respir Cell Mol Biol*. 2011; 44:787–793. [PubMed: 20656951]
31. Elder MJ, Webster SJ, Williams DL, Gaston JS, Goodall JC, et al. TSLP production by dendritic cells is modulated by IL-1 $\beta$  and components of the endoplasmic reticulum stress response. *Eur J Immunol*. 2016; 46:455–463. [PubMed: 26573878]
32. Tsilingiri K, Fornasa G, Rescigno M. Thymic Stromal Lymphopoietin: To Cut a Long Story Short. *Cell Mol Gastroenterol Hepatol*. 2017; 3:174–182. [PubMed: 28275684]
33. Poposki JA, Klingler AI, Stevens WW, Peters AT, Hulse KE, et al. Proprotein convertases generate a highly functional heterodimeric form of thymic stromal lymphopoietin in humans. *J Allergy Clin Immunol*. 2016; 139:1559–1567. [PubMed: 27744031]
34. Nagarkar DR, Poposki JA, Tan BK, Comeau MR, Peters AT, et al. Thymic stromal lymphopoietin activity is increased in nasal polyps of patients with chronic rhinosinusitis. *J Allergy Clin Immunol*. 2013; 132:593–600. [PubMed: 23688414]
35. Kim BS, Siracusa MC, Saenz SA, Noti M, Monticelli LA, et al. TSLP elicits IL-33-independent innate lymphoid cell responses to promote skin inflammation. *Sci Transl Med*. 2013; 5:170ra16.
36. Mjösberg J, Bernink J, Golebski K, Karrich JJ, Peters CP, et al. The transcription factor GATA3 is essential for the function of human type 2 innate lymphoid cells. *Immunity*. 2012; 37:649–659. [PubMed: 23063330]
37. Liu S, Verma M, Michalec L, Liu W, Sripada A, et al. Steroid resistance of airway type 2 innate lymphoid cells from patients with severe asthma: The role of thymic stromal lymphopoietin. *J Allergy Clin Immunol*. 2017; S0091-6749:30660–30667.
38. Kabata H, Moro K, Fukunaga K, Suzuki Y, Miyata J, et al. Thymic stromal lymphopoietin induces corticosteroid resistance in natural helper cells during airway inflammation. *Nat Commun*. 2013; 4:2675. [PubMed: 24157859]
39. Gauvreau GM, O'Byrne PM, Boulet LP, Wang Y, Cockcroft D, et al. Effects of an anti-TSLP antibody on allergen-induced asthmatic responses. *N Engl J Med*. 2014; 370:2102–2110. [PubMed: 24846652]
40. Satoh-Takayama N, Lesjean-Pottier S, Vieira P, Sawa S, Eberl G, et al. IL-7 and IL-15 independently program the differentiation of intestinal CD3(-)NKp46(+) cell subsets from Id2-dependent precursors. *J Exp Med*. 2010; 207:273–280. [PubMed: 20142427]

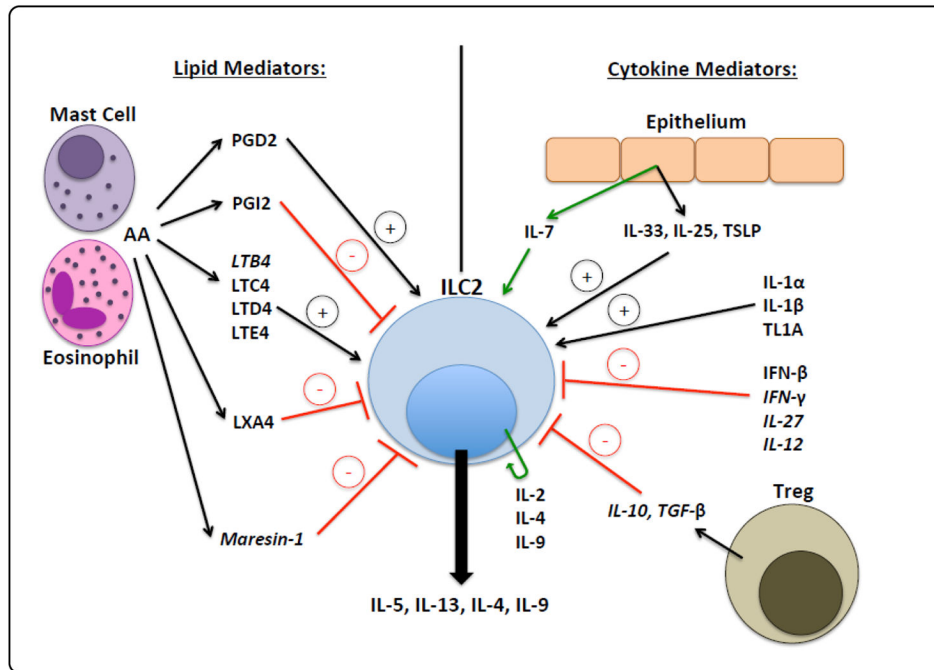


41. Halim TY, Krauss RH, Sun AC, Takei F. Lung natural helper cells are a critical source of Th2 cell-type cytokines in protease allergen-induced airway inflammation. *Immunity*. 2012; 36:451–463. [PubMed: 22425247]
42. Roediger B, Kyle R, Yip KH, Sumaria N, Guy TV, et al. Cutaneous immunosurveillance and regulation of inflammation by group 2 innate lymphoid cells. *Nat Immunol*. 2013; 14:564–573. [PubMed: 23603794]
43. Bal SM, Bernink JH, Nagasawa M, Groot J, Shikhagaie MM, et al. IL-1 $\beta$ , IL-4 and IL-12 control the fate of group 2 innate lymphoid cells in human airway inflammation in the lungs. *Nat Immunol*. 2016; 17:636–645. [PubMed: 27111145]
44. Ohne Y, Silver JS, Thompson-Snipes L, Collet MA. IL-1 is a critical regulator of group 2 innate lymphoid cell function and plasticity. *Nat Immunol*. 2016; 17:646–655. [PubMed: 27111142]
45. Baumann R, Rabaszowski M, Stenin I, Tilgner L, Scheckenbach K, et al. Comparison of the nasal release of IL-4, IL-10, IL-17, CCL13/MCP-4, and CCL26/eotaxin-3 in allergic rhinitis during season and after allergen challenge. *Am J Rhinol Allergy*. 2013; 27:266–272. [PubMed: 23883806]
46. Gour N, Wills-Karp M. IL-4 and IL-13 signaling in allergic airway disease. *Cytokine*. 2015; 75:68–78. [PubMed: 26070934]
47. Doherty TA, Khorram N, Chang JE, Kim HK, Rosenthal P, et al. STAT6 regulates natural helper cell proliferation during lung inflammation initiated by *Alternaria*. *Am J Physiol Lung Cell Mol Physiol*. 2012; 303:L577–588. [PubMed: 22865552]
48. Motomura Y, Morita H, Moro K, Nakae S, Artis D, et al. Basophil-derived interleukin-4 controls the function of natural helper cells, a member of ILC2s, in lung inflammation. *Immunity*. 2014; 40:758–771. [PubMed: 24837103]
49. Darveaux J, Busse WW. Biologics in asthma--the next step toward personalized treatment. *J Allergy Clin Immunol Pract*. 2015; 3:152–160. [PubMed: 25754716]
50. Kim AS, Doherty TA. New and emerging therapies for asthma. *Ann Allergy Asthma Immunol*. 2016; 116:14–17. [PubMed: 26707770]
51. Meylan F, Hawley ET, Barron L, Barlow JL, Penumetcha P, et al. The TNF-family cytokine TL1A promotes allergic immunopathology through group 2 innate lymphoid cells. *Mucosal Immunol*. 2014; 7:958–68. [PubMed: 24368564]
52. Yu X, Pappu R, Ramirez-Carrozzi V, Ota N, Caplazi P, et al. TNF superfamily member TL1A elicits type 2 innate lymphoid cells at mucosal barriers. *Mucosal Immunol*. 2014; 7:730–740. [PubMed: 24220298]
53. Wilhelm C, Hirota K, Stieglitz B, Van Snick J, Tolaini M, et al. An IL-9 fate reporter demonstrates the induction of an innate IL-9 response in lung inflammation. *Nat Immunol*. 2011; 12:1071–1077. [PubMed: 21983833]
54. Turner JE, Morrison PJ, Wilhelm C, Wilson M, Ahlfors H, et al. IL-9-mediated survival of type 2 innate lymphoid cells promotes damage control in helminth-induced lung inflammation. *J Exp Med*. 2013; 210:2951–2965. [PubMed: 24249111]
55. Moro K, Kabata H, Tanabe M, Koga S, Takeno N, et al. Interferon and IL-27 antagonize the function of group 2 innate lymphoid cells and type 2 innate immune responses. *Nat Immunol*. 2016; 17:76–86. [PubMed: 26595888]
56. Duerr CU, McCarthy CD, Mindt BC, Rubio M, Meli AP, et al. Type I interferon restricts type 2 immunopathology through the regulation of group 2 innate lymphoid cells. *Nat Immunol*. 2016; 17:65–75. [PubMed: 26595887]
57. Rigas D, Lewis G, Aron JL, Wang B, Banie H, et al. Type 2 innate lymphoid cell suppression by regulatory T cells attenuates airway hyperreactivity and requires inducible T-cell costimulator-inducible T-cell costimulator ligand interaction. *J Allergy Clin Immunol*. 2017; 139:1468–1477. [PubMed: 27717665]
58. Doherty TA, Broide DH. Pathways to limit group 2 innate lymphoid cell activation. *J Allergy Clin Immunol*. 2017; 139:1465–1467. [PubMed: 28087226]
59. Boyce JA. Mast cells and eicosanoid mediators: a system of reciprocal paracrine and autocrine regulation. *Immunol Rev*. 2007; 217:168–185. [PubMed: 17498059]



60. Davidson AB, Lee TH, Scanlon PD, Solway J, McFadden ER Jr, et al. Bronchoconstrictor effects of leukotriene E4 in normal and asthmatic subjects. *Am Rev Respir Dis.* 1987; 135:333–337. [PubMed: 3028218]
61. Barrett NA, Rahman OM, Fernandez JM, Parsons MW, Xing W, et al. Dectin-2 mediates Th2 immunity through the generation of cysteinyl leukotrienes. *J Exp Med.* 2011; 208:593–604. [PubMed: 21357742]
62. Kim DC, Hsu FI, Barrett NA, Friend DS, Grenningloh R, et al. Cysteinyl leukotrienes regulate Th2 cell-dependent pulmonary inflammation. *J Immunol.* 2006; 176:4440–4448. [PubMed: 16547282]
63. Xue L, Barrow A, Fleming VM, Hunter MG, Ogg G, et al. Leukotriene E4 activates human Th2 cells for exaggerated proinflammatory cytokine production in response to prostaglandin D2. *J Immunol.* 2012; 188:694–702. [PubMed: 22174450]
64. Bankova LG, Lai J, Yoshimoto E, Boyce JA, Austen KF, et al. Leukotriene E4 elicits respiratory epithelial cell mucin release through the G-protein-coupled receptor, GPR99. *Proc Natl Acad Sci U S A.* 2016; 113:6242–6247. [PubMed: 27185938]
65. Bartemes KR, Iijima K, Kobayashi T, Kephart GM, McKenzie AN, et al. IL-33-responsive lineage-CD25+ CD44(hi) lymphoid cells mediate innate type 2 immunity and allergic inflammation in the lungs. *J Immunol.* 2012; 188:1503–1513. [PubMed: 22198948]
66. Lund SJ, Portillo A, Cavagnero K, Baum RE, Naji LH, et al. Leukotriene C4 Potentiates IL-33-Induced Group 2 Innate Lymphoid Cell Activation and Lung Inflammation. *J Immunol.* 2017; 199:1096–1104. [PubMed: 28667163]
67. Xue L, Salimi M, Panse I, Mjösberg JM, McKenzie AN, et al. Prostaglandin D2 activates group 2 innate lymphoid cells through chemoattractant receptor-homologous molecule expressed on TH2 cells. *J Allergy Clin Immunol.* 2014; 133:1184–1194. [PubMed: 24388011]
68. Salimi M, Stöger L, Liu W, Go S, Pavord I, et al. Cysteinyl leukotriene E4 activates human group 2 innate lymphoid cells and enhances the effect of prostaglandin D2 and epithelial cytokines. *J Allergy Clin Immunol.* 2017; S0091-6749:30051–30059.
69. Yokomizo T, Kato K, Hagiya H, Izumi T, Shimizu T, et al. Hydroxyeicosanoids bind to and activate the low affinity leukotriene B4 receptor, BLT2. *J Biol Chem.* 2001; 276:12454–12459. [PubMed: 11278893]
70. Konya V, Mjösberg J. Lipid mediators as regulators of human ILC2 function in allergic diseases. *Immunol Lett.* 2016; 179:36–42. [PubMed: 27396531]
71. Chang JE, Doherty TA, Baum R, Broide D. Prostaglandin D2 regulates human type 2 innate lymphoid cell chemotaxis. *J Allergy Clin Immunol.* 2014; 133:899–901. e3. [PubMed: 24210841]
72. Barnig C, Cernadas M, Dutile S, Liu X, Perrella MA, et al. Lipoxin A4 regulates natural killer cell and type 2 innate lymphoid cell activation in asthma. *Sci Transl Med.* 2013; 5:174ra26.
73. Wojno ED, Monticelli LA, Tran SV, Alenghat T, Osborne LC, et al. The prostaglandin D(2) receptor CRTH2 regulates accumulation of group 2 innate lymphoid cells in the inflamed lung. *Mucosal Immunol.* 2015; 8:1313–23. [PubMed: 25850654]
74. Kim SH, Kim YK, Park HW, Jee YK, Bahn JW, et al. Association between polymorphisms in prostanoid receptor genes and aspirin-intolerant asthma. *Pharmacogenet Genomics.* 2007; 17:295–304. [PubMed: 17496729]
75. Schulman ES, Newball HH, Demers LM, Fitzpatrick FA, Adkinson NF Jr. Anaphylactic release of thromboxane A2, prostaglandin D2, and prostacyclin from human lung parenchyma. *Am Rev Respir Dis.* 1981; 124:402–406. [PubMed: 6170242]
76. Claar D, Hartert TV, Peebles RS Jr. The role of prostaglandins in allergic lung inflammation and asthma. *Expert Rev Respir Med.* 2015; 9:55–72. [PubMed: 25541289]
77. Fanning LB, Boyce JA. Lipid mediators and allergic diseases. *Ann Allergy Asthma Immunol.* 2013; 111:155–162. [PubMed: 23987187]
78. Dorris SL, Peebles RS Jr. PGI2 as a regulator of inflammatory diseases. *Mediators Inflamm.* 2012; 2012:926968. [PubMed: 22851816]
79. Zhou W, Toki S, Zhang J, Goleniewska K, Newcomb DC, et al. Prostaglandin I2 Signaling and Inhibition of Group 2 Innate Lymphoid Cell Responses. *Am J Respir Crit Care Med.* 2016; 193:31–42. [PubMed: 26378386]

80. Christie PE, Spur BW, Lee TH. The effects of lipoxin A4 on airway responses in asthmatic subjects. *Am Rev Respir Dis*. 1992; 145:1281–1284. [PubMed: 1595991]
81. Krishnamoorthy N, Burkett PR, Dalli J, Abdulnour RE, Colas R, et al. Cutting edge: maresin-1 engages regulatory T cells to limit type 2 innate lymphoid cell activation and promote resolution of lung inflammation. *J Immunol*. 2015; 194:863–7. [PubMed: 25539814]
82. Walford HH, Doherty TA. Diagnosis and management of eosinophilic asthma: a US perspective. *J Asthma Allergy*. 2014; 7:53–65. [PubMed: 24748808]
83. Chen R, Smith SG, Salter B, El-Gammal A, Oliveria JP, et al. Allergen-induced Increases in Sputum Levels of Group 2 Innate Lymphoid Cells in Asthmatic Subjects. *Am J Respir Crit Care Med*. 2017
84. Wheatley LM, Togias A. Clinical practice. Allergic rhinitis. *N Engl J Med*. 2015; 372:456–463. [PubMed: 25629743]
85. Asaka D, Yoshikawa M, Nakayama T, Yoshimura T, Moriyama H, et al. Elevated levels of interleukin-33 in the nasal secretions of patients with allergic rhinitis. *Int Arch Allergy Immunol*. 2012; 158:47–50. [PubMed: 22627366]
86. Fan D, Wang X, Wang M, Wang y, Zhang L, et al. Allergen-Dependent Differences in ILC2s Frequencies in Patients With Allergic Rhinitis. *Allergy Asthma Immunol Res*. 2016; 8:216–22. [PubMed: 26922931]
87. Cahill KN, Bensko JC, Boyce JA, Laidlaw TM. Prostaglandin D2: A dominant mediator of aspirin exacerbated respiratory disease. *J Allergy Clin Immunol*. 2015; 135:245–252. [PubMed: 25218285]
88. Liu T, Kanaoka Y, Barrett NA, Feng C, Garofalo D, et al. Aspirin-Exacerbated Respiratory Disease Involves a Cysteinyl Leukotriene-Driven IL-33-Mediated Mast Cell Activation Pathway. *J Immunol*. 2015; 195:3537–45. [PubMed: 26342029]
89. Buchheit KM, Cahill KN, Katz HR, Murphy KC, Feng C, et al. Thymic stromal lymphopoietin controls prostaglandin D generation in patients with aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol*. 2016; 137:1566–1576. [PubMed: 26691435]

**Figure 1.**

Lipids and cytokines that regulate ILC2s. Mediators only found in mouse are italicized and green lines represent maintenance/homeostatic cytokines.